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THE EFFECTS OF EXERCISE ON DENDRITIC SPINE DENSITY: IMPLICATIONS FOR EXERCISE-INDUCED MEMORY ENHANCEMENT

EGZERSİZİN DENDRİTİK DİKENLERİN YOĞUNLUĞU ÜZERİNE ETKİLERİ: EGZERSİZE BAĞLI BELLEK GELİŞTİRME UYGULAMALARI

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Abstract

The purpose of this brief review was to highlight the potential mediational role of dendritic spine morphology on the exercise-memory interaction. I first start out delineating the role of dendritic spine density on episodic memory function and then discuss mechanisms involved in spine density alteration. Following this, I discuss the effects that exercise has on dendritic spine density, including its underlying mechanisms. Ultimately, this discussion will provide us with greater insights on the mediating mechanisms through which exercise may influence episodic memory function.

Keywords: BDNF; cognition; long-term potentiation; synapse

Özet

Bu kısa derlemenin amacı, dendritik dikenlerin morfolojisinin egzersiz--bellek etkileşimi üzerine etkisini incelemektir. Bu bağlamda, öncelikle dendritik dikenlerin yoğunluğunun epizodik bellek üzerine etkisinin altı çizilecek, ardından dikenlerin yoğunluğundaki değişiklikler mekanizması ile ilgili tartışılacaktır. Bundan sonrasında, egzersizin altta yatan mekanizmaları ile birlikte dendritik dikenlerin yoğunluğu üzerindeki etkisi tartışılacaktır. Son olarak, tartışma egzersizin epizodik bellek kapasitesi üzerindeki aracı rolü hakkındaki bilgilerimizi arttırmamıza imkan sağlayacaktır.

Anahtar Kelimeler: BDNF; biliş; uzun dönemli potansiyasyon; sinaps

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1. Introduction

Both acute and chronic exercise have been shown to enhance episodic memory (Frith, Sng, & Loprinzi, 2017; Haynes Iv, Frith, Sng, & Loprinzi, 2018; Loprinzi, Scott, Ikuta, Addoh, & Tucker, 2018; Sng, Frith, & Loprinzi, 2018), which is the retrieval of retrospective information from a spatio-temporal context (Tulving, 1983). We have previously discussed various mechanisms through which both acute and chronic exercise may subserve episodic memory (Loprinzi, 2018; Loprinzi, Edwards, & Frith, 2017; Loprinzi & Frith, 2018; Loprinzi, Ponce, & Frith, 2018). These include, for example, exercise-induced alterations in neurotrophic factors, hormones, and long-term potentiation. The present review extends this body of work by highlighting the role of exercise on synaptic plasticity, and specifically, dendritic spine density. This is of critical importance, as the brain's ability to retain information depends on the strength of existing synapses and on the appearance or elimination of dendritic spines (Fregozo & Vega, 2012; Kasai, Fukuda, Watanabe, Hayashi-Takagi, & Noguchi, 2010).

I first start out delineating the role of dendritic spine density on episodic memory function and then discuss mechanisms involved in spine density alteration. Following this, I discuss the effects that exercise has on dendritic spine density, including its underlying mechanisms. Ultimately, this discussion will provide us with greater insights on the mediating mechanisms through which exercise may influence episodic memory function.

2. Dendritic Spines and Memory Function

As illustrated elsewhere (Poo et al., 2016), structural and molecular mediators of memory are complex, and unlikely attributed to a single structure or protein. Of interest to this paper, however, is the role of dendritic spines and how they are formed and modified during learning. A spine consists of three basic components, including a base structure joined to the dendrite, a neck, and a head which may connect with an axon. Normal dendritic spine density ranges from 0.2 to 3.5 spines per 1 μm of dendrite (Benavides-Piccione, Feraud-Espinosa, Robles, Yuste, & DeFelipe, 2013). Further, spines are often classified into three morphological groups, including, thin, stubby, and mushroom types. Spines may become stable for days to years (Zuo, Lin, Chang, & Gan, 2005), and the amount of stable spines correlates with memory performance (Yang, Pan, & Gan, 2009). Notably, however, the actin filaments that supports the spine turns over in minutes to hours, with 80% of F-actin in spines turning over every minute (Star, Kwiatkowski, & Murthy, 2002).

Dendritic spines exist in certain types of neurons, such as pyramidal neurons in the cortex, hippocampal neurons and Purkinje cells in the cerebellum (Kasai, Fukuda, Watanabe, Hayashi-Takagi, & Noguchi, 2010). Each neuron may include over 10,000 spines, each representing a point of synaptic contact (Kasai, Fukuda, Watanabe, Hayashi-Takagi, & Noguchi, 2010). Relatedly, dendritic spines are sites for synaptic communication, and

thus, alterations of the spine (via LTP and LTD (Trommald, Hulleberg, & Andersen, 1996)) play an important role in synaptic plasticity, and ultimately, memory function. Notably, spines with large postsynaptic densities have more AMPA receptors (Noguchi et al., 2011), which may facilitate LTP. Further, spine neck size may also influence NMDA-dependent Ca^{2+} signaling (Noguchi, Matsuzaki, Ellis-Davies, & Kasai, 2005).

In addition to increasing the surface area for synaptic contact, the majority of dendritic spines have an excitatory synapse, with Ca^{2+} concentrated in the spine, as well as voltage-gated Na channels in the spine (Rose, Kovalchuk, Eilers, & Konnerth, 1999), suggesting that dendritic spines play a critical role in the formation and plasticity of functional neural networks involved in memory function (Yuste, 2011). Thus, structural changes in dendrites may be an important mechanism of long-term information storage. This has been demonstrated in empirical work showing that spine loss among networks that were active during learning impairs long-term memory function (Sanders, Cowansage, Baumgartel, & Mayford, 2012). Further, stress-induction has been shown to impair memory and reduce the density of CA3 dendritic spines, and blocking this stress-induced effect prevents spine loss and restores memory function (Chen et al., 2010). Other work also demonstrates that genetic manipulation of spine turnover influences storage capacity and memory function (Frank et al., 2018). Certain neuropsychiatric conditions, such as depression (decreased number of spines), fragile X syndrome (elongated, tortuous spines), and Down syndrome (fewer spines, some with large heads), have spine- and memory-related impairments (McCann & Ross, 2017).

3. Mechanisms Involved in Dendritic Spine Density

Several key reviews have provided mechanistic insight on the regulation of dendritic spines (Basu & Lamprecht, 2018; Borovac, Bosch, & Okamoto, 2018; Cornelia Koeberle et al., 2017; Ebrahimi & Okabe, 2014; Fregozo & Vega, 2012; Hotulainen et al., 2009; Kumar et al., 2016; Merriam et al., 2013; Penzes & Rafalovich, 2012; Sala & Segal, 2014; Spence & Soderling, 2015; Sutton & Schuman, 2006). Remodeling of the actin cytoskeleton is a likely candidate behind the structural alterations of dendritic spines, and plays a fundamental role in the formation, elimination, motility, stability, size, and shape of the spine (Penzes & Rafalovich, 2012).

In neurons, actin consists of soluble monomeric G-actin and polymerized F-actin filaments. Polymerization of free G-actin is regulated by various pathways, activated by select surface receptors, such as NMDA receptors (Cingolani & Goda, 2008). Upon activation of this NMDA receptor, the dendritic spine undergoes a temporary increase in Ca^{2+} (Sobczyk & Svoboda, 2007), which then activates calcium-sensing calmodulin, ultimately activating various kinases (e.g., CamKI, CamKII, and CamKIV) (Hook & Means, 2001). These kinases then proceed on to phosphorylate targets involved in spine

structural plasticity (e.g., Kalirin-7) (Penzes & Rafalovich, 2012).

More specifically, this process can be conceptualized as a 4-step process, including an initial basal state, F-actin disassembly, F-actin assembly, and F-actin stabilization (Borovac, Bosch, & Okamoto, 2018). During the basal state, bundled F-actin maintains a stable spine structure. During the F-actin disassembly stage, kinases (e.g., CamKII) are activated and become detached from F-actin and then unbundle the filaments. Cofilin/ADF then enters the spine and severs the filaments. Drebrin and α -actinin reduce the concentration of filaments in the spine. Aip1 and Arp2/3 enter the spine and branch F-actin together, resulting in enlargement of spine size. Following this enlargement, stabilizing proteins, such as Drebrin, α -actinin, and inactivated CamKII return to their basal state, bind to F-actin, re-bundle and cross-link the reorganized filaments, ultimately stabilizing the enlarged dendritic spine (Borovac, Bosch, & Okamoto, 2018).

Activation of 5-HT_{2A} receptors in pyramidal neurons has been shown to increase spine size via Kalirin-7-Rac1-PAK-dependent mechanisms (Jones et al., 2009). Data also supports the role of dopamine (Solis, Limon, Flores-Hernandez, & Flores, 2007; Wang & Deutch, 2008) and the cholinergic system (Sherren & Pappas, 2005) in regulating spine morphology. Relatedly, numerous studies have reported BDNF-induced changes in spine morphology (Lu, Christian, & Lu, 2008), with TrkB-deficient mice having fewer dendritic spines in the CA1 hippocampal neurons (Luikart & Parada, 2006).

4. Effects of Exercise on Dendritic Spine Density

Treadmill exercise (two weeks of daily exercise) in mice has been shown to attenuate stress-induced dendritic spine elimination (Chen et al., 2017). This stress-induced memory impairment is attenuated with exercise, mediated via neurogenesis and dendritic remodeling (Yau et al., 2011). Similarly, exercise has been shown to attenuate diabetes-induced dendritic spine elimination, likely through BDNF augmentation (Stranahan et al., 2009). Other work also supports this exercise-induced attenuation of spine elimination in Parkinson's disease (Toy et al., 2014) and multiple sclerosis (Rossi et al., 2009).

5. Mechanisms through which Exercise Influences Dendritic Spine Density

Dendritic spines may respond morphologically to a large variety of physiological stimuli, including acute and chronic exercise. As we thoroughly discussed elsewhere (Loprinzi, Ponce, & Frith, 2018), skeletal muscle contraction, via exercise, may activate peripheral afferent muscle spindle fibers, which have direct projections to the brainstem, and ultimately, the hippocampus. Similarly, exercise-induced lung expansion and heart rate increases will activate afferent vagus nerve fibers, which also have projections to the hippocampus. Such exercise-induced neuronal

excitability may increase various neurotransmitters (e.g., glutamate) to initiate dendritic spine morphology. As discussed above, key proteins, such as BDNF, and surface receptors, such as NMDA, play an important role in spine alterations.

Previous research by Dietrich et al. (Dietrich et al., 2005) demonstrated that the level of phosphorylation of NR1 and NR2 subunits of the rat cerebral cortex NMDA receptor was upregulated with exercise. Further, the NMDA receptor channel open rate also increased with exercise. Molteni et al. (Molteni, Ying, & Gomez-Pinilla, 2002) also demonstrated that exercise increased the expression of NR1, NR2A, and NR2B mRNA in the rat hippocampus after 3 and 7 days of exercise. Exercise has also been shown to increase BDNF levels (Loprinzi & Frith, 2018) and BDNF may help upregulate the function of the NMDA receptor (Caldeira et al., 2007; Clarke & Johnson, 2008; Kim et al., 2012).

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References

- Basu, S., & Lamprecht, R. (2018). The Role of Actin Cytoskeleton in Dendritic Spines in the Maintenance of Long-Term Memory. *Front Mol Neurosci*, 11, 143. doi: 10.3389/fnmol.2018.00143
- Benavides-Piccione, R., Feraud-Espinosa, I., Robles, V., Yuste, R., & DeFelipe, J. (2013). Age-based comparison of human dendritic spine structure using complete three-dimensional reconstructions. *Cereb Cortex*, 23(8), 1798-1810. doi: 10.1093/cercor/bhs154
- Borovac, J., Bosch, M., & Okamoto, K. (2018). Regulation of actin dynamics during structural plasticity of dendritic spines: Signaling messengers and actin-binding proteins. *Mol Cell Neurosci*, 91, 122-130. doi: 10.1016/j.mcn.2018.07.001
- Caldeira, M. V., Melo, C. V., Pereira, D. B., Carvalho, R. F., Carvalho, A. L., & Duarte, J. B. (2007). BDNF regulates the expression and traffic of NMDA receptors in cultured hippocampal neurons. *Mol Cell Neurosci*, 35(2), 208-219. doi: 10.1016/j.mcn.2007.02.019
- Chen, K., Zhang, L., Tan, M., Lai, C. S., Li, A., Ren, C., & So, K. F. (2017). Treadmill exercise suppressed stress-induced dendritic spine elimination in mouse barrel cortex and improved working memory via BDNF/TrkB pathway. *Transl Psychiatry*, 7(3), e1069. doi: 10.1038/tp.2017.41
- Chen, Y., Rex, C. S., Rice, C. J., Dube, C. M., Gall, C. M., Lynch, G., & Baran, T. Z. (2010). Correlated memory defects and hippocampal dendritic spine loss after acute stress involve corticotropin-releasing hormone signaling. *Proc Natl Acad Sci U S A*, 107(29), 13123-13128. doi: 10.1073/pnas.1003825107
- Cingolani, L. A., & Goda, Y. (2008). Actin in action: the interplay between the actin cytoskeleton and synaptic efficacy. *Nat Rev Neurosci*, 9(5), 344-356. doi: 10.1038/nrn2373
- Clarke, R. J., & Johnson, J. W. (2008). Voltage-dependent gating of NR1/2B NMDA receptors. *J Physiol*, 586(23), 5727-5741. doi: 10.1113/jphysiol.2008.160622
- Cornelia Koeberle, S., Tanaka, S., Kuriu, T., Iwasaki, H., Koeberle, A., Schulz, A., . . . Okabe, S. (2017). Developmental stage-dependent regulation of spine formation by calcium-calmodulin-dependent protein kinase II α and Rap1. *Sci Rep*, 7(1), 13409. doi: 10.1038/s41598-017-13728-y
- Dietrich, M. O., Mantese, C. E., Porciuncula, L. O., Ghisleni, G., Vinade, L., Souza, D. O., & Portela, L. V. (2005). Exercise affects glutamate receptors in postsynaptic densities from cortical mice brain. *Brain Res*, 1065(1-2), 20-25. doi: 10.1016/j.brainres.2005.09.038
- Ebrahimi, S., & Okabe, S. (2014). Structural dynamics of dendritic spines: molecular composition, geometry and functional regulation. *Biochim Biophys Acta*, 1838(10), 2391-2398. doi: 10.1016/j.bbamm.2014.06.002
- Frank, A. C., Huang, S., Zhou, M., Gdalyahu, A., Kastellakis, G., Silva, T. K., . . . Silva, A. J. (2018). Hotspots of dendritic spine turnover facilitate

clustered spine addition and learning and memory. *Nat Commun*, 9(1), 422. doi: 10.1038/s41467-017-02751-2

Fregozo, C. S., & Vega, M. I. (2012). Actin-binding proteins and signalling pathways associated with the formation and maintenance of dendritic spines. *Neurologia*, 27(7), 421-431.

Frith, E., Sng, E., & Loprinzi, P. D. (2017). Randomized controlled trial evaluating the temporal effects of high-intensity exercise on learning, short-term and long-term memory, and prospective memory. *Eur J Neurosci*, 46(10), 2557-2564. doi: 10.1111/ejn.13719

Haynes Iv, J. T., Frith, E., Sng, E., & Loprinzi, P. D. (2018). Experimental Effects of Acute Exercise on Episodic Memory Function: Considerations for the Timing of Exercise. *Psychol Rep*, 33294118786688. doi: 10.1177/0033294118786688

Hook, S. S., & Means, A. R. (2001). Ca(2+)/CaM-dependent kinases: from activation to function. *Annu Rev Pharmacol Toxicol*, 41, 471-505. doi: 10.1146/annurev.pharmtox.41.1.471

Hotulainen, P., Llano, O., Smirnov, S., Tanhuanpaa, K., Faix, J., Rivera, C., & Lappalainen, P. (2009). Defining mechanisms of actin polymerization and depolymerization during dendritic spine morphogenesis. *J Cell Biol*, 185(2), 323-339. doi: 10.1083/jcb.200809046

Jones, K. A., Srivastava, D. P., Allen, J. A., Strachan, R. T., Roth, B. L., & Penzes, P. (2009). Rapid modulation of spine morphology by the 5-HT_{2A} serotonin receptor through kalirin-7 signaling. *Proc Natl Acad Sci U S A*, 106(46), 19575-19580. doi: 10.1073/pnas.0905884106

Kasai, H., Fukuda, M., Watanabe, S., Hayashi-Takagi, A., & Noguchi, J. (2010). Structural dynamics of dendritic spines in memory and cognition. *Trends Neurosci*, 33(3), 121-129. doi: 10.1016/j.tins.2010.01.001

Kim, J. H., Roberts, D. S., Hu, Y., Lau, G. C., Brooks-Kayal, A. R., Farb, D. H., & Russek, S. J. (2012). Brain-derived neurotrophic factor uses CREB and Egr3 to regulate NMDA receptor levels in cortical neurons. *J Neurochem*, 120(2), 210-219. doi: 10.1111/j.1471-4159.2011.07555.x

Kumar, A., Paeger, L., Kosmas, K., Kloppenburg, P., Noegel, A. A., & Peche, V. S. (2016). Neuronal Actin Dynamics, Spine Density and Neuronal Dendritic Complexity Are Regulated by CAP2. *Front Cell Neurosci*, 10, 180. doi: 10.3389/fncel.2016.00180

Loprinzi, P. D. (2018). IGF-1 in exercise-induced enhancement of episodic memory. *Acta Physiol (Oxf)*, e13154. doi: 10.1111/apha.13154

Loprinzi, P. D., Edwards, M. K., & Frith, E. (2017). Potential avenues for exercise to activate episodic memory-related pathways: a narrative review. *Eur J Neurosci*, 46(5), 2067-2077. doi: 10.1111/ejn.13644

Loprinzi, P. D., & Frith, E. (2018). A brief primer on the mediational role of BDNF in the exercise-memory link. *Clin Physiol Funct Imaging*. doi: 10.1111/cpf.12522

Loprinzi, P. D., Ponce, P., & Frith, E. (2018). Hypothesized mechanisms through which acute exercise influences episodic memory. *Physiol Int*, 105(4), 285-297. doi: 10.1556/2060.105.2018.4.28

Loprinzi, P. D., Scott, T. M., Ikuta, T., Addoh, O., & Tucker, K. L. (2018). Association of physical activity on changes in cognitive function: Boston Puerto Rican Health Study. *Phys Sportsmed*, 1-5. doi: 10.1080/00913847.2018.1547087

Lu, Y., Christian, K., & Lu, B. (2008). BDNF: a key regulator for protein synthesis-dependent LTP and long-term memory? *Neurobiol Learn Mem*, 89(3), 312-323. doi: 10.1016/j.nlm.2007.08.018

Luikart, B. W., & Parada, L. F. (2006). Receptor tyrosine kinase B-mediated excitatory synaptogenesis. *Prog Brain Res*, 157, 15-24.

McCann, R. F., & Ross, D. A. (2017). A Fragile Balance: Dendritic Spines, Learning, and Memory. *Biol Psychiatry*, 82(2), e11-e13. doi: 10.1016/j.biopsych.2017.05.020

Merriam, E. B., Millette, M., Lombard, D. C., Saengsawang, W., Fothergill, T., Hu, X., . . . Dent, E. W. (2013). Synaptic regulation of microtubule dynamics in dendritic spines by calcium, F-actin, and drebrin. *J Neurosci*, 33(42), 16471-16482. doi: 10.1523/JNEUROSCI.0661-13.2013

Molteni, R., Ying, Z., & Gomez-Pinilla, F. (2002). Differential effects of acute and chronic exercise on plasticity-related genes in the rat hippocampus revealed by microarray. *Eur J Neurosci*, 16(6), 1107-1116.

Noguchi, J., Matsuzaki, M., Ellis-Davies, G. C., & Kasai, H. (2005). Spine-neck geometry determines NMDA receptor-dependent Ca²⁺ signaling in dendrites. *Neuron*, 46(4), 609-622. doi: 10.1016/j.neuron.2005.03.015

Noguchi, J., Nagaoka, A., Watanabe, S., Ellis-Davies, G. C., Kitamura, K., Kano, M., . . . Kasai, H. (2011). In vivo two-photon uncaging of glutamate revealing the structure-function relationships of dendritic spines in the neocortex of adult mice. *J Physiol*, 589(Pt 10), 2447-2457. doi: 10.1111/jphysiol.2011.207100

Penzes, P., & Rafalovich, I. (2012). Regulation of the actin cytoskeleton in dendritic spines. *Adv Exp Med Biol*, 970, 81-95. doi: 10.1007/978-3-

7091-0932-8_4

Poo, M. M., Pignatelli, M., Ryan, T. J., Tonegawa, S., Bonhoeffer, T., Martin, K. C., . . . Stevens, C. (2016). What is memory? The present state of the engram. *BMC Biol*, 14, 40. doi: 10.1186/s12915-016-0261-6

Rose, C. R., Kovalchuk, Y., Eilers, J., & Konnerth, A. (1999). Two-photon Na⁺ imaging in spines and fine dendrites of central neurons. *Pflügers Arch*, 439(1-2), 201-207.

Rossi, S., Furlan, R., De Chiara, V., Musella, A., Lo Giudice, T., Mataluni, G., . . . Centonze, D. (2009). Exercise attenuates the clinical, synaptic and dendritic abnormalities of experimental autoimmune encephalomyelitis. *Neurobiol Dis*, 36(1), 51-59. doi: 10.1016/j.nbd.2009.06.013

Sala, C., & Segal, M. (2014). Dendritic spines: the locus of structural and functional plasticity. *Physiol Rev*, 94(1), 141-188. doi: 10.1152/physrev.00012.2013

Sanders, J., Cowansage, K., Baumgartel, K., & Mayford, M. (2012). Elimination of dendritic spines with long-term memory is specific to active circuits. *J Neurosci*, 32(36), 12570-12578. doi: 10.1523/JNEUROSCI.1131-12.2012

Sherren, N., & Pappas, B. A. (2005). Selective acetylcholine and dopamine lesions in neonatal rats produce distinct patterns of cortical dendritic atrophy in adulthood. *Neuroscience*, 136(2), 445-456. doi: 10.1016/j.neuroscience.2005.08.053

Sng, E., Frith, E., & Loprinzi, P. D. (2018). Temporal Effects of Acute Walking Exercise on Learning and Memory Function. *Am J Health Promot*, 32(7), 1518-1525. doi: 10.1177/0890117117749476

Sobczyk, A., & Svoboda, K. (2007). Activity-dependent plasticity of the NMDA-receptor fractional Ca²⁺ current. *Neuron*, 53(1), 17-24. doi: 10.1016/j.neuron.2006.11.016

Solis, O., Limon, D. I., Flores-Hernandez, J., & Flores, G. (2007). Alterations in dendritic morphology of the prefrontal cortical and striatum neurons in the unilateral 6-OHDA-rat model of Parkinson's disease. *Synapse*, 61(6), 450-458. doi: 10.1002/syn.20381

Spence, E. F., & Soderling, S. H. (2015). Actin Out: Regulation of the Synaptic Cytoskeleton. *J Biol Chem*, 290(48), 28613-28622. doi: 10.1074/jbc.R115.655118

Star, E. N., Kwiatkowski, D. J., & Murthy, V. N. (2002). Rapid turnover of actin in dendritic spines and its regulation by activity. *Nat Neurosci*, 5(3), 239-246. doi: 10.1038/nn811

Stranahan, A. M., Lee, K., Martin, B., Maudsley, S., Golden, E., Cutler, R. G., & Mattson, M. P. (2009). Voluntary exercise and caloric restriction enhance hippocampal dendritic spine density and BDNF levels in diabetic mice. *Hippocampus*, 19(10), 951-961. doi: 10.1002/hipo.20577

Sutton, M. A., & Schuman, E. M. (2006). Dendritic protein synthesis, synaptic plasticity, and memory. *Cell*, 127(1), 49-58. doi: 10.1016/j.cell.2006.09.014

Toy, W. A., Petzinger, G. M., Leyshon, B. J., Akopian, G. K., Walsh, J. P., Hoffman, M. V., . . . Jakowec, M. W. (2014). Treadmill exercise reverses dendritic spine loss in direct and indirect striatal medium spiny neurons in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) mouse model of Parkinson's disease. *Neurobiol Dis*, 63, 201-209. doi: 10.1016/j.nbd.2013.11.017

Trommald, M., Hulleberg, G., & Andersen, P. (1996). Long-term potentiation is associated with new excitatory spine synapses on rat dentate granule cells. *Learn Mem*, 3(2-3), 218-228.

Tulving, E. (1983). Elements of episodic memory: Oxford University Press.

Wang, H. D., & Deutch, A. Y. (2008). Dopamine depletion of the prefrontal cortex induces dendritic spine loss: reversal by atypical antipsychotic drug treatment. *Neuropsychopharmacology*, 33(6), 1276-1286. doi: 10.1038/sj.npp.1301521

Yang, G., Pan, F., & Gan, W. B. (2009). Stably maintained dendritic spines are associated with lifelong memories. *Nature*, 462(7275), 920-924. doi: 10.1038/nature08577

Yau, S. Y., Lau, B. W., Tong, J. B., Wong, R., Ching, Y. P., Qiu, G., . . . So, K. F. (2011). Hippocampal neurogenesis and dendritic plasticity support running-improved spatial learning and depression-like behaviour in stressed rats. *PLoS One*, 6(9), e24263. doi: 10.1371/journal.pone.0024263

Yuste, R. (2011). Dendritic spines and distributed circuits. *Neuron*, 71(5), 772-781. doi: 10.1016/j.neuron.2011.07.024

Zuo, Y., Lin, A., Chang, P., & Gan, W. B. (2005). Development of long-term dendritic spine stability in diverse regions of cerebral cortex. *Neuron*, 46(2), 181-189. doi: 10.1016/j.neuron.2005.04.001